

**Genetic Analysis of Anxiety
Related Behaviors by Gene
Chip and *In situ*
Hybridization of the
Hippocampus and
Amygdala of C57BL/6J and
AJ Mice Brains**

INTRODUCTION

To study the relationship between an animal's behavior, the brain systems involved, and the underlying genetic make-up, a researcher must move from the behavior and gross anatomy down to the molecular level. Initial tests performed by researchers Chantal Mathis, Steven Paul, Jacqueline Crawley, and Howard Gershenfeld have characterized the inbred strains of A/J and C57BL/6J mice as being on the extremes of an emotional behavioral continuum. Tests with A/J and C57BL/6J mice show the levels of exploratory behavior in light/dark models are much higher for C57BL/6J mice than for A/J mice. Exploratory activity in an open field also shows C57BL/6J as having significantly higher levels of exploratory behavior than A/J. Passive avoidance performance show no evidence of acquisition of passive avoidance task at low levels of footshock in C57BL/6J mice, yet A/J mice readily acquire memory task at low levels of footshock.

C57BL/6J mice are

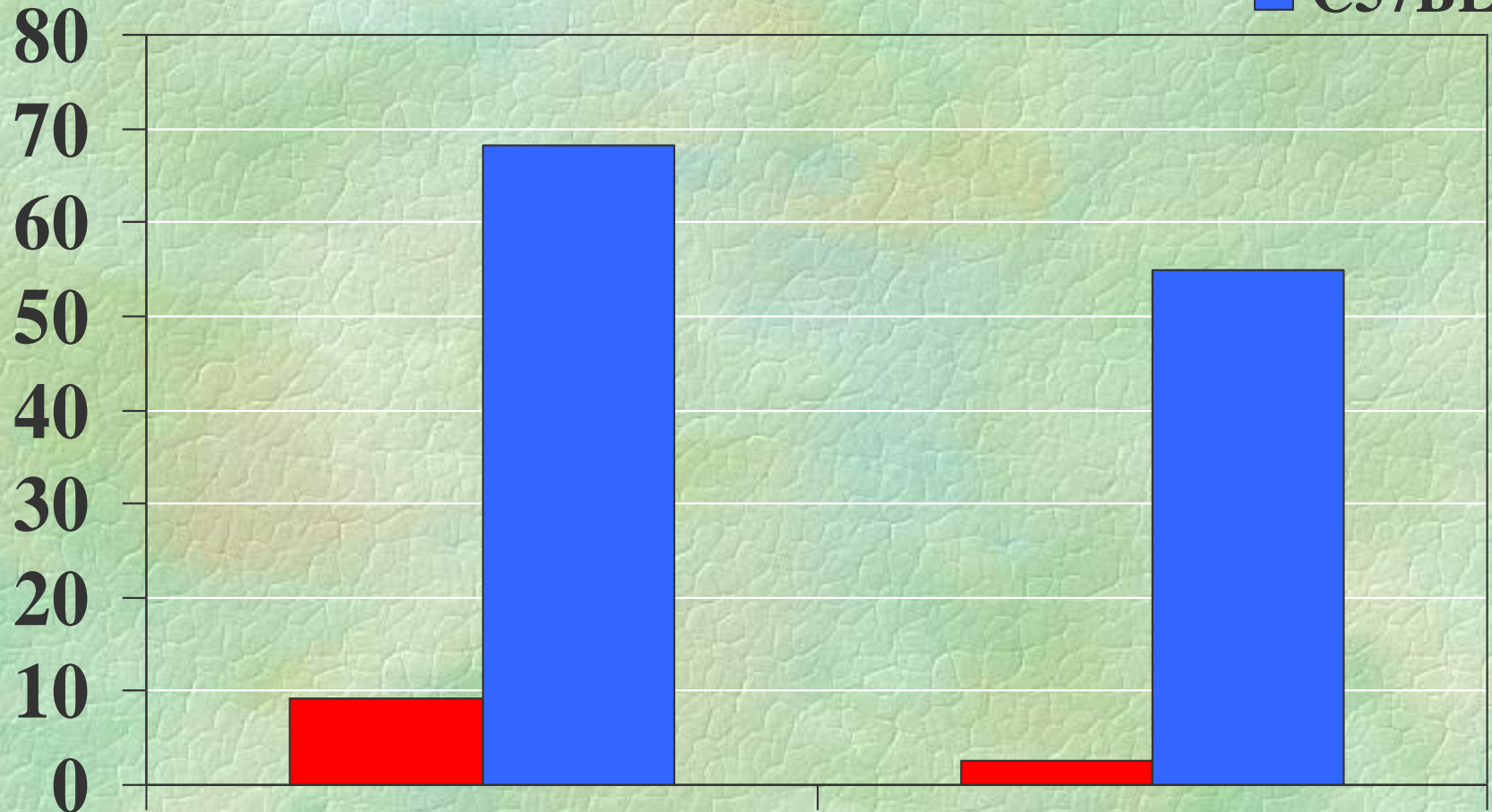
- less "emotional" and "behaviorally inhibited"
- more active
- less sensitive to novelty
than A/J mice.

These characteristics allow one to characterize A/J mice as inhibited and C57BL/6J mice as less inhibited. These significant differences between the phenotypes of these two inbred stains gave indirect evidence that genes were contributing to the variation in phenotype and gene mapping could then be conducted for quantitative trait loci (QTL) analysis.

Behavioral Strain Differences

■ A/J

■ C57BL6/J



LD transitions

Rearing

Mouse Strain

METHODS

Traditional methods in molecular biology generally work on a "one gene in one experiment" basis, which means that the throughput is very limited and the "whole picture" of gene function is hard to obtain. In the past several years, a new technology, called a "GeneChip", DNA microarray or microarray assay has been developed. Researchers have used GeneChip probe arrays to study the regulation of gene expression associated with a wide variety of basic biological functions, including development, hormonal signaling, and circadian rhythms. An experiment with a single GeneChip can provide information on thousands of genes simultaneously - a dramatic increase in throughput. This technology monitors the whole genome on a single chip so a better picture of the simultaneous interactions among thousands of genes is seen and determinations of expression level (abundance) of genes can be made.



Gene Chip



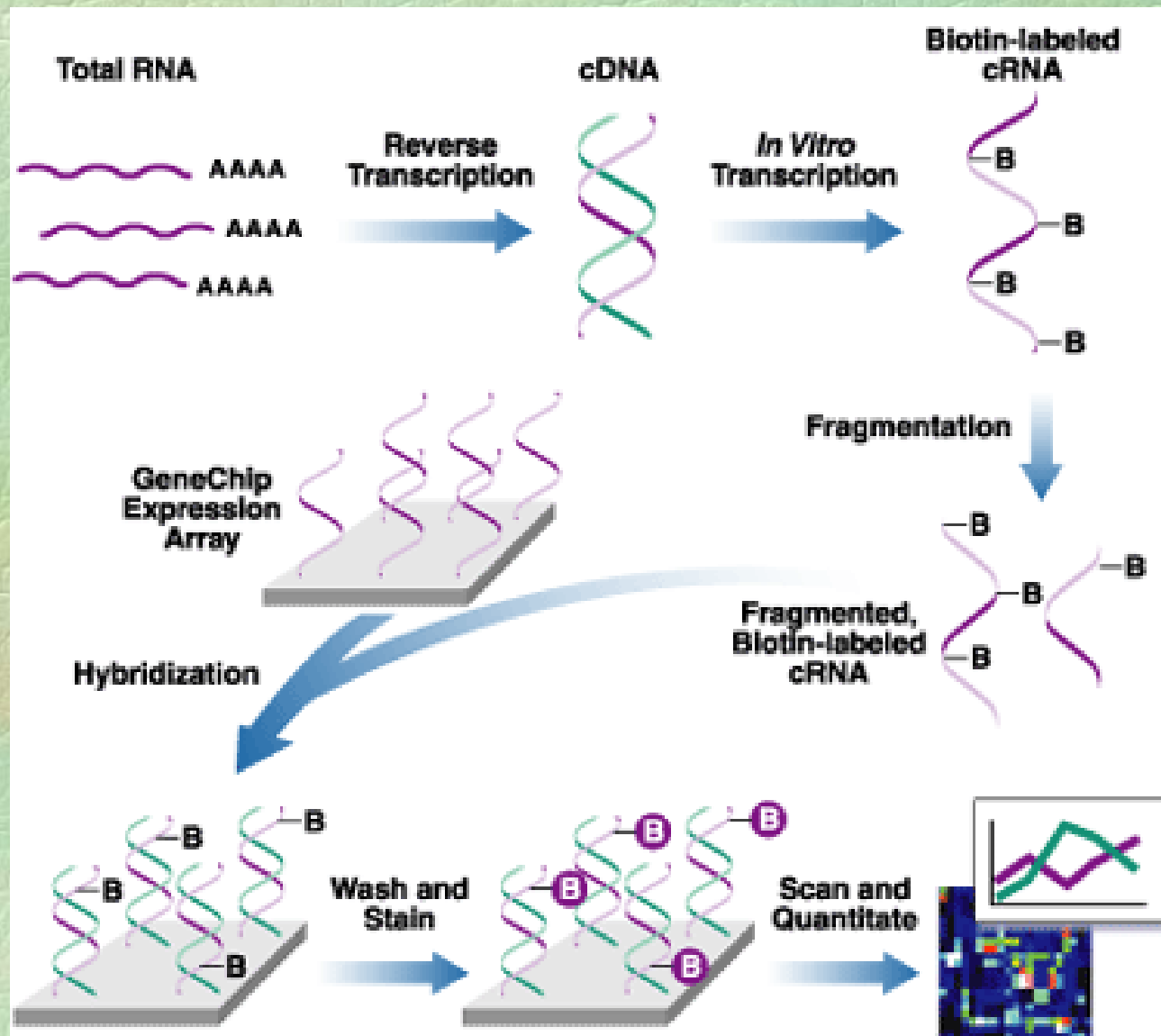
**Relative size of
Gene Chip**

Pictures modified from GeneDetect.com

GeneChip Analysis of Hippocampus from A/J and C57BL/6J Mice

The Gershenfeld Lab used Gene Chips to determine which genes are activated and which genes are repressed when comparing the hippocampus of A/J and C57BL/6J mice.

1. Obtain the cells of the hippocampus of A/J and C57BL/6J mice. Extract the mRNA because DNA microarray analysis is based on the assumption that transcriptions level of genes are equal to the mRNA present in cell.
2. Since mRNA degrades so easily, make cDNA from mRNA using reverse transcription. Degrade the remaining mRNA.
3. Label cDNA of the A/J samples and C57BL/6J samples with florescent labels.
4. Each spot on gene chip slide is made of DNA, (actually a 25 mer oligonubcleotide) that can base pair w/ cDNA.
5. Hybridize cDNA samples to chip and incubate. Each chip covers 7000 genes. Base-pairing (i.e., A-T and G-C for DNA; A-U and G-C for RNA) or hybridization is the underlining principle of DNA microarray.
6. Wash unbound cDNA off. All that is left is what has specifically bound to the DNA spots.
7. Scan the chip with laser to detect the bound cDNA. Images from the A/J and C57BL/6J chips are stored in computer for later signal processing and comparisons. Sophisticated software algorithms compare expression levels of each gene and calculate the extent (fold) increases, decreases, or no significant changes.



Gene Chip Protocol (modified from GeneDetect.com)

***In Situ* Analysis of Hippocampus from A/J and C57BL/6J Mice**

In order to validate the GeneChip data obtained and to find the patterns of genes and their expression levels in the hippocampus and amygdala of the brains of the A/J and C57BL/6J mice, in situ hybridization will be used. In situ hybridization allows a particular mRNA species to be localized in its normal anatomical environment (i.e., tissue section.)

1. Since the data on the GeneChip experiment suggests that genes RGS5, YWHAQ, and EBAF are those involved in "anxiety behavior," 45 bp oligonucleotides of complimentary sequences to genes RGS5, YWHAQ, and EBAF are obtained.
2. The oligo probes will be labeled radioactively by incorporating about 16 bp worth of ³⁵Sulphur onto them.
3. These labeled oligos will be hybridized to slides of 14um sections of hippocampus and amygdala tissue sections from A/J and C57BL/6J mice. Unbound oligos will be washed off to avoid background.
4. Visualization of the gene expression is done by exposing the slides to photographic film for 4-7 days, and then developing the film.

Brain cross section showing
fluorescently labeled probe

(modified from
www.affymetrix.com)

PROBE TO COMMON SEQUENCE



SPLICE VARIANT-SPECIFIC PROBE



RESULTS

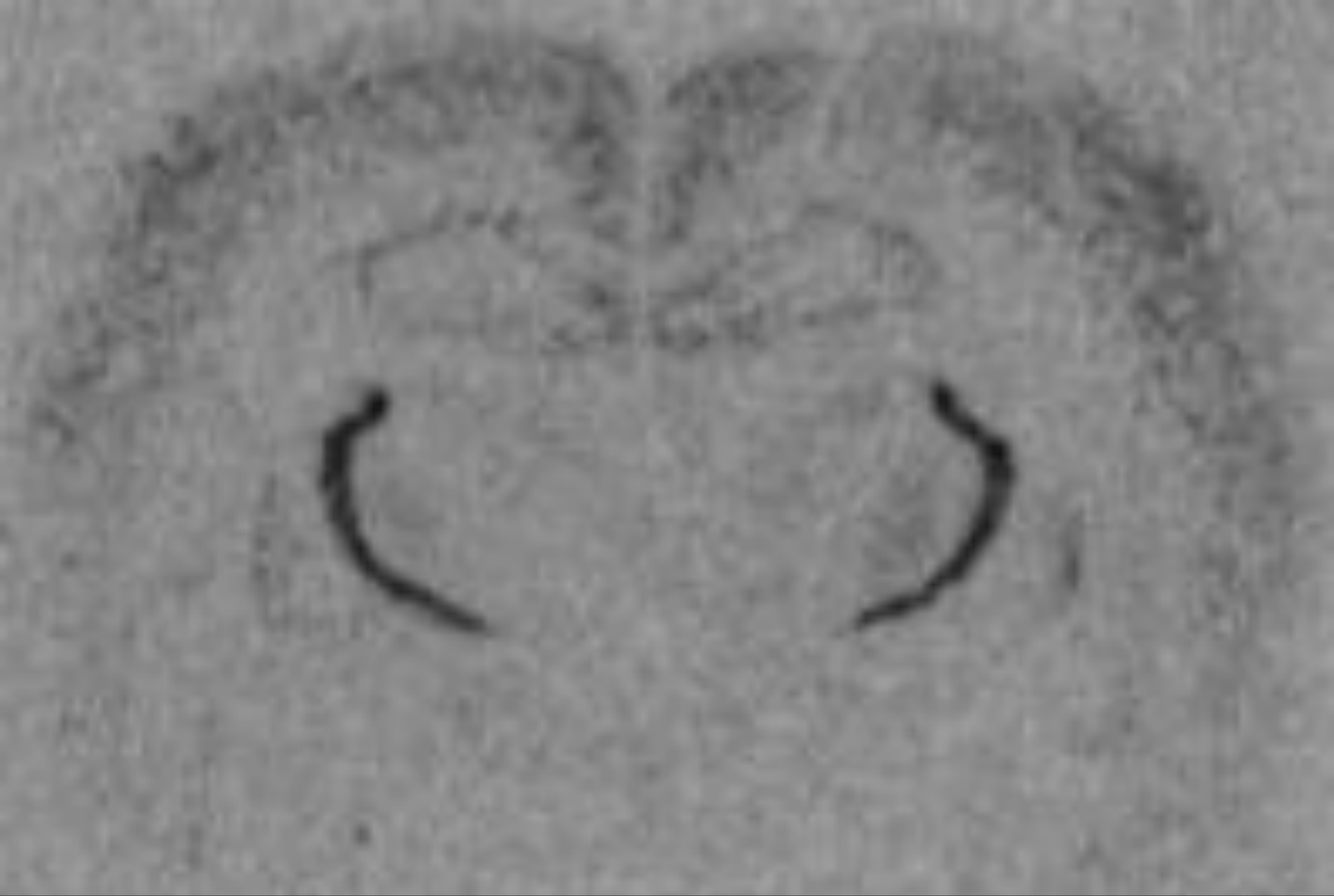
GeneChip analysis show increased expressions of genes RGS5, YWHAQ, and EBAF. This suggests that these are the genes involved in "anxiety behavior" in A/J and C57BL/6J mice.

In situ hybridization will be performed to compare the level and/or location difference of RGS 5, YWHAQ, and EBAF expression in the brain regions of A/J and C57BL/6J mice. Ideally, there will be more RGS 5, YWHAQ, and EBAF expressed in the hypothalamus of C57BL/6J than that in A/J. Real time PCR will be used to further validate the potential difference.



in Situ hybridization showing mice hippocampus

(modified from www.unifr.ch/biochem/ABRECHT/LECTURES/InSitu)



in Situ Hybridization showing mice hippocampus

(modified from www.unifr.ch/biochem/ABRECHT/LECTURES/InSitu)

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LITERATURE CITED

- Blumstein, L. K. and J. N. Crawley (1983). "Further characterization of a simple, automated exploratory model for the anxiolytic effects of benzodiazepines." Pharmacology, Biochemistry & Behavior **18**(1): 37-40.
- Crawley, J. and F. K. Goodwin (1980). "Preliminary report of a simple animal behavior model for the anxiolytic effects of benzodiazepines." Pharmacology, Biochemistry & Behavior **13**(2): 167-70.
- Crawley, J. N. (1981). "Neuropharmacologic specificity of a simple animal model for the behavioral actions of benzodiazepines." Pharmacology, Biochemistry & Behavior **15**(5): 695-9.
- Crawley, J. N. and L. G. Davis (1982). "Baseline exploratory activity predicts anxiolytic responsiveness to diazepam in five mouse strains." Brain Research Bulletin **8**(6): 609-12.
- Gershenfeld, H. K. and S. M. Paul (1998). "Towards a genetics of anxious temperament: from mice to men." Acta Psychiatr Scand Suppl **393**: 56-65.
- Mathis, C., P. E. Neumann, et al. (1995). "Analysis of Anxiety-related behaviors and responses to benzodiazepine-related drugs in AxB and BxA Recombinant Inbred Mouse Strains." Behavior Genetics **25**(6): 557-568.
- Mathis, C., S. M. Paul, et al. (1994). "Characterization of benzodiazepine-sensitive behaviors in the A/J and C57BL/6J inbred strains of mice." Behavior Genetics **24**(2): 171-80.